

FORM PTO-1390 (Modified)  
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U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

## TRANSMITTAL LETTER TO THE UNITED STATES

235.00310101

DESIGNATED/ELECTED OFFICE (DO/EO/US)

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

CONCERNING A FILING UNDER 35 U.S.C. 371

Unassigned **10/019642**INTERNATIONAL APPLICATION NO.  
**PCT/US00/18051**INTERNATIONAL FILING DATE  
**30/06/2000**PRIORITY DATE CLAIMED  
**01/07/1999**

## TITLE OF INVENTION

**FERTILITY IMPAIRING VACCINE CONTAINING AVIAN ZONA PELLUCIDA PROTEIN AND METHOD OF USE**

APPLICANT(S) FOR DO/EO/US

**FAYRER-HOSKEN et al.**

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☐ This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.
4. ☒ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
  - a. ☐ is attached hereto (required only if not communicated by the International Bureau).
  - b. ☐ has been communicated by the International Bureau.
  - c. ☒ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
  - a. ☐ is attached hereto.
  - b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
  - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
  - b. ☐ have been communicated by the International Bureau.
  - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
  - d. ☒ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☒ A copy of the International Search Report (PCT/ISA/210).

**Items 13 to 20 below concern document(s) or information included:**

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☐ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
20. ☐ A second copy of the published international application under 35 U.S.C. 154(d)(4).
21. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
22. ☒ Certificate of Mailing by Express Mail
23. ☒ Other items or information:

**Itemized Return-Stamped postcard; Transmittal Letter (1 p.); and check in the amount of \$920.00 for filing fee.**

28 DEC 2001

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.101) <b>Unassigned</b>	INTERNATIONAL APPLICATION NO. <b>PCT/US00/18051</b>	ATTORNEY'S DOCKET NUMBER <b>235.00310101</b>
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24. The following fees are submitted:				<b>CALCULATIONS PTO USE ONLY</b>	
<b>BASIC NATIONAL FEE ( 37 CFR 1.492 (a) (1) - (5) ) :</b>					
<input type="checkbox"/>	Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO .....		<b>\$1040.00</b>		
<input checked="" type="checkbox"/>	International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO .....		<b>\$890.00</b>		
<input type="checkbox"/>	International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO .....		<b>\$740.00</b>		
<input type="checkbox"/>	International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) .....		<b>\$710.00</b>		
<input type="checkbox"/>	International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) .....		<b>\$100.00</b>		
<b>ENTER APPROPRIATE BASIC FEE AMOUNT =</b>				<b>\$890.00</b>	
Surcharge of <b>\$130.00</b> for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492 (e)). <input type="checkbox"/> 20 <input checked="" type="checkbox"/> 30				<b>\$130.00</b>	
<b>CLAIMS</b>	<b>NUMBER FILED</b>	<b>NUMBER EXTRA</b>	<b>RATE</b>		
Total claims	50 - 20 =	30	x \$18.00	<b>\$540.00</b>	
Independent claims	2 - 3 =	0	x \$84.00	<b>\$0.00</b>	
Multiple Dependent Claims (check if applicable).			<input checked="" type="checkbox"/>	<b>\$280.00</b>	
<b>TOTAL OF ABOVE CALCULATIONS =</b>				<b>\$1,840.00</b>	
<input checked="" type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				<b>\$920.00</b>	
<b>SUBTOTAL =</b>				<b>\$920.00</b>	
Processing fee of <b>\$130.00</b> for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.492 (f)). <input type="checkbox"/> 20 <input type="checkbox"/> 30				<b>\$0.00</b>	
<b>TOTAL NATIONAL FEE =</b>				<b>\$920.00</b>	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). <input type="checkbox"/>				<b>\$0.00</b>	
<b>TOTAL FEES ENCLOSED =</b>				<b>\$920.00</b>	
				<b>Amount to be refunded</b>	\$
				<b>charged</b>	\$

- a. ☒ A check in the amount of **920.00** to cover the above fees is enclosed.
- b. ☐ Please charge my Deposit Account No. \_\_\_\_\_ in the amount of \_\_\_\_\_ to cover the above fees. A duplicate copy of this sheet is enclosed.
- c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. **13-4895**. A duplicate copy of this sheet is enclosed.
- d. ☐ Fees are to be charged to a credit card. **WARNING: Information on this form may become public. Credit card information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

**NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.**

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43,022

REGISTRATION NUMBER

*December 28, 2001*  
DATE

**Application Data Sheet**

**Application Information**

Application Number::

Filing Date::

Application Type:: Regular

Subject Matter:: Utility

Suggested Classification::

Suggested Group Art Unit::

CD-ROM or CD-R?::

Number of CD Disks::

Number of Copies of CDs::

Sequence Submission::

Computer Readable Form (CRF)?::

Number of Copies of CRF::

Title:: FERTILITY IMPAIRING VACCINE  
CONTAINING AVIAN ZONA PELLUCIDA  
PROTEIN AND METHOD OF USE

Attorney Docket Number:: 235.00310101

Request for Early Publication?:: No

Request for Non-Publication?:: No

Suggested Drawing Figure::

Total Drawing Sheets:: 0

Small Entity?:: Yes

Latin Name::

Variety Denomination Name::

Petition Included?:: No

Petition Type::

Licensed US Govt. Agency:: No

Contract or Grant Numbers::

Secrecy Order in Parent Appl.?:: No

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Representative Customer Number::	26813	
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**OR**

Representative Designation::	Registration Number::	Representative Name::

**Domestic Priority Information**

Application::	Continuity Type::	Parent Application::	Parent Filing Date::
This Application	National Stage of	PCT/US00/18051	06/30/00
PCT/US00/18051	Non-Provisional of	60/141,929	07/01/99
PCT/US00/18051	Non-Provisional of	60/162,984	11/02/99

**Foreign Priority Information**

Country::	Application Number::	Filing Date::	Priority Claimed::

**Assignee Information**

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Country of Mailing Address::

Postal or Zip Code of Mailing Address::

5      **FERTILITY IMPAIRING VACCINE CONTAINING AVIAN ZONA**  
         **PELLUCIDA PROTEIN AND METHOD OF USE**

10      This application claims the benefit of U.S. Provisional Applications  
Serial No. 60/141,929, filed July 1, 1999, and 60/162,984, filed November 2,  
1999.

**Background of the Invention**

15      Population control of both domesticated and wild, free-roaming  
animals has been a major problem around the world. Overpopulation in dogs and  
cats results in millions of euthanasias of unwanted pets yearly in the United  
States alone, and overpopulation of wild animals causes destruction of habitat  
and threatens other species within the ecosystem. In humane shelters population  
control of unwanted pets is currently achieved through euthanasia of the  
animals. In general, after capture, dogs are held for a period of one week. If  
20      they are not adopted, they are humanely destroyed.

         Finding a socially acceptable as well as a safe and effective means  
of controlling excess populations of animals has been extremely difficult.  
Traditional methods of population control in dogs have been largely  
unsuccessful. Surgical spaying is a laborious procedure, requiring the initial  
25      induction of the animal, gas anesthesia during surgery, a surgical pack with  
suture materials and post-operative medications. Common surgical  
complications include problems associated with the procedure itself, allergic  
reactions to anesthetics or post-operative medications, and adverse local or  
systemic effects during the recovery period. Examples include ovarian remnant  
30      syndrome, where dogs continue to cycle despite being spayed, uterine infections,  
abdominal hemorrhage, and premature opening of the suture line. A substantial  
recovery period is typically needed even after an uncomplicated procedure.

Surgical spaying is also expensive, and pet owners are often unwilling to assume the costs.

5 The use of a reversible contraceptive is an attractive alternative to surgical spaying. However, conventional hormonal therapies that have been used to curb pet overpopulation are inconvenient because they usually require daily administration of the drug. Furthermore, most protracted hormonal therapies have undesirable side effects such as uterine infections, mammary cancer, and diabetes.

10 A vaccine comprising porcine zona pellucida protein (pZP) and an adjuvant comprising synthetic trehalose dicorynomycolate has been successfully used to cause immunocontraception in horses (P. Willis et al., J. Equine Vet. Sci. 14:364-370 (1994)) and elephants (R. Fayer-Hosken, Wildlife Soc. Bull., 25(1):18-21 (1997)). Injection of total heat solubilized pZP into horses results in an immunocontraceptive effect that lasts for about 12 months and has been effective in reducing fertility of free-roaming horses (Kirkpatrick et al., 15 J. Reprod. Fert. 94:437-444 (1992)). The pZP vaccine has been used for more than 8 years in horses with no adverse reactions; nor does the vaccine affect the feti of pregnant mares.

20 A pZP vaccine has been administered to dogs as well, albeit with somewhat less success. Mahi-Brown et al. have shown that infertility in the female dog can be achieved by vaccinating with a crude pZP preparation, but vaccination was accompanied by abnormal estrus cycles and other deleterious side effects, such as ovarian cyst formation (J. Exp. Zool., 222:89-95 (1982); Am. J. Reprod. Immunol. Microbiol., 18: 94-103 (1988)).

25 Porcine zona pellucida protein is obtained from pig ovaries, thus the amount of vaccine that can be prepared is limited by the supply of pig ovaries. Dunbar et al. (e.g., EP 599822, U.S. Pat. No. 5,637,300) have experimented with reproductive control in non-rodent mammals using a recombinant zona pellucida protein. However, due to limitations imposed by 30 recombinant DNA technology and available expression systems, the recombinant protein lacks the glycosylation pattern of the native glycoprotein and its immunogenicity has not been satisfactorily demonstrated. What is clearly needed to combat overpopulation of domesticated and wild, free-roaming



animals is a low cost contraceptive or sterilant that possesses a high degree of efficiency, lacks of harmful side effects, is amenable to remote delivery, requires a minimal number of administrations, and is easy to produce.

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### **Summary of the Invention**

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The present invention provides a fertility impairing vaccine and a method for impairing fertility in an animal. One embodiment of the fertility impairing vaccine, referred to herein as a protein vaccine, comprises a polypeptide comprising at least one avian zona pellucida protein or an immunogenic fragment thereof. These proteins are related to the mammalian zona pellucida proteins. Optionally, the protein vaccine of the invention further includes a polypeptide comprising a porcine zona pellucida protein or an immunogenic fragment thereof.

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Another embodiment of the fertility impairing vaccine, referred to herein as a polynucleotide vaccine, comprises a polynucleotide comprising at least one nucleotide sequence that encodes a polypeptide comprising an avian zona pellucida protein or an immunogenic fragment thereof. Optionally, the polynucleotide vaccine of the invention further includes a polynucleotide comprising a nucleotide sequence that encodes a polypeptide comprising a porcine zona pellucida protein or an immunogenic fragment thereof.

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The fertility impairing vaccine of the invention optionally further includes an immunological adjuvant or immunostimulant, preferably synthetic trehalose dicorynomycolate (STDCM). Also optionally, the fertility impairing vaccine contains an oil, preferably squalene oil.

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The method for impairing fertility in the animal comprises administering to the animal the fertility impairing vaccine of the invention in a manner and an amount effective to cause fertility impairment in the animal. When administered as an immunocontraceptive, the fertility impairing vaccine causes temporary, reversible infertility in the animal. When administered as an immunosterilant, the fertility impairing vaccine causes permanent, irreversible infertility in the animal. In dogs and cats, for example, immunosterilization is

far preferable to surgical sterilization and hormone regimens as a population control tool.

The fertility impairing vaccine of the invention can generally be used for immunological control of fertility and reproduction in any desired animal population, whether domestic or wild. As an example, the fertility impairing vaccine can be used to control ferrel dog and cat populations by development of a species-specific oral delivery vehicle. As another example, the fertility impairing vaccine can be used to control wild horse, deer or elephant populations when formulated as a single dose dart or bullet that can be administered without capture or confinement of the animal.

#### **Detailed Description of the Preferred Embodiments**

The fertility impairing vaccine of the invention contains a polypeptide comprising at least one avian zona pellucida protein (aZP) or an immunogenic fragment thereof, or a polynucleotide having a nucleotide sequence encoding a polypeptide comprising an avian zona pellucida protein or an immunogenic fragment thereof. The avian zona pellucida protein is preferably, but need not be, substantially pure. The avian zona pellucida protein or immunogenic fragment thereof can be a naturally occurring protein, a chemically or enzymatically synthesized protein, or a recombinant protein. The avian zona pellucida protein or immunogenic fragment thereof is preferably, but need not be, glycosylated. In a glycosylated avian zona pellucida protein (i.e., an avian zona pellucida glycoprotein), the glycosylation pattern is preferably equivalent to the glycosylation pattern found on a native (i.e., naturally occurring) glycoprotein. Naturally occurring avian zona pellucida protein is preferably obtained from a chicken. Avian zona pellucida protein can be conveniently obtained from bird ovaries or from an egg cell of the bird and/or the surrounding extracellular matrix and tissue, particularly the perivitelline membrane, for example from an oocyte or unfertilized egg of a bird at any stage of development.

In a preferred embodiment of the fertility impairing vaccine, the avian zona pellucida protein used in the vaccine or encoded by the

polynucleotide used in the vaccine is a total avian zona pellucida protein. A total avian zona pellucida protein preparation includes a plurality of immunoreactive proteins, some of which have been identified in chickens as gp42 and gp97 (Y. Takeuchi et al., Eur. J. Biochem. 260:736-742 (1999)), and gp34 and gp95 (M. Wacławek et al., Biol. Reprod. 59:1230-1239 (1998)). Example I, below, identifies immunoreactive chicken proteins having molecular weights of 70 kD, 40 kD and 35 kD. The fertility impairing vaccine can, alternatively, contain fewer than all the avian zona pellucida proteins; for example it can contain the 40 kD and the 35 kD proteins but not the 70 kD protein.

Purity of the avian zona pellucida protein or glycoprotein can be evaluated analytically using a combination or series of two-dimensional polyacrylamide gel electrophoresis, sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) with both silver staining and Coomassie Blue staining, and Western blot analysis, as described in the following Examples. Glycoproteins typically migrate electrophoretically in gels as broad smears rather than narrow bands, as a result of the variable levels of negative charge present in the constituent oligosaccharide chains. A total avian zona pellucida protein preparation isolated from bird eggs typically migrates as three distinct smears in the gel electrophoretic experiments, and shows the immunological reactivity in Western blot analysis using a polyclonal antibody raised in rabbits to highly purified total porcine zona pellucida protein. In a substantially pure avian zona pellucida protein preparation, no contaminating proteins having electromigration patterns different from those exhibited by the avian zona pellucida proteins are detectable in the two-dimensional gel, the SDS-PAGE gel, or Western blot analyses.

An immunogenic fragment of an avian zona pellucida protein or glycoprotein is a peptide fragment, preferably a glycosylated peptide fragment, that elicits an immune response in a subject to which it is administered. An immune response includes either or both of a cellular immune response or production of antibodies, and can include activation of the subject's B cells, T cells, helper T cells or other cells of the subject's immune system. For example,

an immune response is evidenced by a detectable anti-aZP antibody level in the subject using ELISA substantially as described in Example III.

Immunogenicity of an avian zona pellucida protein fragment can be determined, for example, by administering the adjuvanted candidate fragment to the subject, then observing of the associated immune response by analyzing anti-aZP fragment antibody titers in serum. An immunogenic peptide fragment preferably contains more than seven amino acids, more preferably at least about 10 amino acids, most preferably at least about 20 amino acids.

In a preferred embodiment of the fertility impairing vaccine, the polypeptide used in the vaccine or encoded by the polynucleotide used in the vaccine further includes at least one epitope, such as a T cell, helper T cell or B cell epitope. The epitopes can be derived from the species to which the vaccine is to be administered, from the species that was the source of the zona pellucida protein or immunogenic fragment thereof, or from any other species, including a virus, bacterium, or parasite. T cell, helper T cell or B cell epitopes or epitope mimics have been identified in zona pellucida proteins (Garza et al., J. Reprod. Immunol., February 1998, pp. 87-101). The use of immune cell epitopes derived from an immunogenic organism, such as a pathogenic parasite, is preferred. For example, the polynucleotide can encode a chimeric peptide comprising a T cell or helper T cell epitope from a parasite and a B cell epitope from a zona pellucida protein, such as a porcine or avian zona pellucida protein (Bagavant et al., Biol. Reprod., March 1997, pp. 764-770). A vaccine for use as an immunosterilant preferably contains a polypeptide that contains at least one T cell epitope (or a polynucleotide functionally encoding such a polypeptide), whereas a vaccine for use as an immunocontraceptive preferably includes a polypeptide that encodes at least one B cell epitope (or a polynucleotide functionally encoding such a polypeptide).

A polynucleotide encoding the polypeptide comprising the avian zona pellucida protein or immunogenic fragment thereof can include DNA, RNA, a modified nucleic acid, or any combination thereof. The polynucleotide can be supplied as part of a vector or as a "naked" polynucleotide. General methods for construction, production and administration of polynucleotide vaccines are known in the art, e.g. F. Vogel et al., *Clin. Microbiol. Rev.* 8:406-

410 (1995). Polynucleotides can be generated by means standard in the art, such as by recombinant techniques, or by enzymatic or chemical synthesis.

A polynucleotide used in a fertility impairing vaccine of the invention is preferably one that functionally encodes an avian zona pellucida protein. A protein is "functionally encoded" if it is capable of being expressed from the genetic construct that contains it. For example, the polynucleotide can include one or more expression control sequences, such as *cis*-acting transcription/translation regulatory sequences, including one or more of the following: a promoter, response element, an initiator sequence, an enhancer, a ribosome binding site, an RNA splice site, an intron element, a polyadenylation site, and a transcriptional terminator sequence, which are operably linked to the coding sequence and are, either alone or in combination, capable of directing expression in the target animal. An expression control sequence is "operably linked" to a coding sequence if it is positioned on the construct such that it does, or can be used to, control or regulate transcription or translation of that coding sequence. Preferred expression control sequences include strong and/or inducible *cis*-acting transcription/translation regulatory sequences such as those derived from metallothionine genes, actin genes, myosin genes, immunoglobulin genes, cytomegalovirus (CMV), SV40, Rous sarcoma virus, adenovirus, bovine papilloma virus, and the like.

The coding and expression control sequences for the avian zona pellucida protein are preferably constructed in a vector, such as a plasmid of bacterial origin, a cosmid, episome, or a viral vector, for administration to a target animal. A vector useful in the present invention can be circular or linear, single-stranded or double stranded. There are numerous plasmids known to those of ordinary skill in the art useful for the production of polynucleotide vaccine plasmids. A specific embodiment employs constructs using the plasmid pcDNA3.1 as the vector (InVitrogen Corporation, Carlsbad, CA). In addition, the vector construct can contain immunostimulatory sequences (ISS) that stimulate the animal's immune system. Other possible additions to the polynucleotide vaccine constructs include nucleotide sequences coding cytokines, such as granulocyte macrophage colony stimulating factor (GM-CSF) or interleukin-12 (IL-12). The cytokines can be used in various combinations to

fine-tune the response of the animal's immune system, including both antibody and cytotoxic T lymphocyte responses, to bring out the specific level of response needed to affect the animal's reproductive system.

Alternatively, the vector can be a viral vector, including an  
5 adenovirus vector, and adenovirus associated vector, or a retroviral vector. Preferably the viral vector is a nonreplicating retroviral vector such as the Moloney murine leukemia virus (N2) backbone as described by Irwin et al. (*J. Virology* 68:5036-5044 (1994)).

Optionally, a fertility impairing protein vaccine of the invention  
10 further includes a polypeptide comprising a porcine zona pellucida protein or an immunogenic fragment thereof. The porcine zona pellucida protein or immunogenic fragment thereof can be a naturally occurring protein, a chemically or enzymatically synthesized protein, or a recombinant protein. The porcine zona pellucida protein or immunogenic fragment thereof is preferably,  
15 but need not be, glycosylated. In a glycosylated porcine zona pellucida protein (i.e., a porcine zona pellucida glycoprotein), the glycosylation pattern is preferably equivalent to the glycosylation pattern found on a native (i.e., naturally occurring) glycoprotein. Immunogenicity of the porcine zona pellucida fragment is as generally described for avian zona pellucida protein.

Without intending to be bound by any particular theory or mechanism, the  
20 porcine zona pellucida protein or immunogenic fragment thereof can be included in the fertility impairing vaccine as an adjuvant to enhance the recipient's immune response, or for its independent and/or synergistic effect in impairing fertility in the recipient. For example, a fertility impairing protein vaccine  
25 substantially comprising chicken zona pellucida protein, which is readily available and relatively inexpensive to isolate, can be augmented by the addition of a small amount of porcine zona pellucida protein, which is more difficult and expensive to obtain. Notwithstanding the above, any desired ratio of avian zona pellucida protein to porcine zona pellucida protein can be used in this  
30 embodiment of the vaccine of the invention. The relative amounts of avian zona pellucida protein and porcine zona pellucida protein used in this embodiment of the vaccine depend on the nature of the animal being vaccinated and the immunogenic response generated in the animal by the vaccine. Preferably the

ratio of avian zona pellucida to porcine zona pellucida (aZP: pZP) used in this embodiment of the vaccine is between 100:1 and 1:100.

Similarly, a polynucleotide vaccine of the invention optionally includes, in addition to a polynucleotide encoding a avian zona pellucida protein or immunogenic fragment thereof, a polynucleotide encoding a porcine zona pellucida protein or an immunogenic fragment thereof. The porcine zona pellucida polynucleotide functionally encodes porcine zona pellucida protein and can take the form of various genetic constructs as generally described for the polynucleotide encoding avian zona pellucida protein. Polynucleotides encoding avian and porcine zona pellucida proteins can constitute part of the same vector or can be delivered to the recipient on different vectors.

The fertility impairing vaccine of the invention, whether it contains a polypeptide (e.g., conjugated or non-conjugated avian zona pellucida protein) or a polynucleotide encoding a polypeptide, optionally further includes an immunological adjuvant to enhance the immunological response of the subject to the immunogenic polypeptide or polynucleotide. Examples of adjuvants include Freund's Complete Adjuvant, Freund's Incomplete Adjuvant, Freund's mycotoxin free adjuvant, aluminum hydroxide, EQUIMUNE (a deproteinized highly purified cell wall extract derived from non-pathogenic *Mycobacteria* spp., Acemannan (a long-chain polydispersed  $\beta(1,4)$  linked mannan polymer interspersed with O-acetylated groups, permulium, and an adjuvant comprising an immunostimulant such as synthetic trehalose dicorynomycolate (STDCM). The vaccine can also optionally include an oil, such as squalene oil, drakeol, or vegetable oil, which can also have an adjuvant effect (see P. Willis et al., J. Equine Vet. Sci., 14, 364-370 (1994)). It should be noted, however, that companion birds exhibit a sensitivity to oil-based adjuvants, which cause necrotic granulomas at the site of administration. An adjuvant comprising synthetic trehalose dicorynemycolate, squalene oil, and a surfactant such as lecithin is preferred for use in mammals. Lecithin typically includes phosphatidyl choline. An adjuvant comprising aluminum hydroxide is preferred for use in birds.

When an oil adjuvant is used, homogenization of the adjuvant, such as Freund's adjuvant, with the aqueous zona pellucida protein or polynucleotide

solution can be accomplished using any convenient means known in the art, such that the oil disperses within the aqueous solution to form an oil in water emulsion. Oil droplet sizes of about 200 nm or less are particularly preferred as they produce a more uniform and stable suspension. A particularly preferred fertility impairing vaccine comprises a predetermined amount of zona pellucida protein or polynucleotide and a pharmaceutically acceptable immunogenic level of adjuvant in an emulsion containing about 10% oil phase and about 90% aqueous phase.

In another embodiment of the invention, the fertility impairing vaccine, whether it contains a conjugated or a non-conjugated avian zona pellucida protein or a polynucleotide encoding said polypeptide, contains no adjuvant; essentially the fertility impairing vaccine of this embodiment is an aqueous avian zona pellucida protein or polynucleotide solution that delivers the intended amount of avian zona pellucida protein or polynucleotide to the recipient.

In a preferred embodiment of the protein vaccine for use in mammals, the vaccine comprises oil, preferably a biodegradable oil such as squalene oil, in an amount of about 2.5% to about 15%, preferably about 8% to about 12%. In preparing the composition it is advantageous to combine a concentrated oily adjuvant composition with an aqueous solution of the immunogen, aZP protein. Typically, the composition is prepared using an adjuvant concentrate which contains lecithin (about 5% to about 15 % wt/vol, preferably about 12% wt /vol) and STDCM (preferably about 25 mg/mL to about 50 mg/mL) in squalene oil. The term % wt/vol means grams per 100 mL of liquid. The aqueous solution containing the isolated aZP protein is typically a phosphate-buffered saline (PBS) solution, and additionally preferably contains Tween 80 (about 0.2% vol/vol to about 0.8% vol/vol, preferably about 0.4% vol/vol). See J.A. Rudbach et al., "Ribi Adjuvants: Chemistry, Biology and Utility in Vaccines for Human and Veterinary Medicine," in The Theory and Practical Application of Adjuvants, D.E.S. Stewart-Tull, Ed., John Wiley & Sons, New York, NY (1995)). Homogenization of the oily adjuvant concentrate with the aqueous aZP solution can be accomplished using any convenient means known in the art, such that the oil disperses within the aqueous solution to form



an oil in water emulsion. Oil droplet sizes of about 200 nm or less are particularly preferred as they produce a more uniform and stable suspension. A particularly preferred vaccine comprises predetermined amounts of aZP and STDCM in an emulsion containing about 10% squalene oil and about 90% aqueous phase.

The fertility impairing vaccine is administered in a manner effective to cause impaired fertility in an animal. An "animal," as that term is used herein, is an oocyte-producing organism that is capable of voluntary mobility. Preferred animals include mammals and birds. More preferably, the animal is a mammal; most preferably it is a deer, horse, elephant, rat, mouse, rabbit, ferret, cat or dog. Birds include but are not limited to free-ranging birds and domesticated birds, including food birds such as chickens, turkeys, and waterfowl, and companion birds in the order Psittaciformes, Passeriformes, Columbiformes, Falconiformes, such as budgerigars, canaries, finches, cockatiels, lovebirds, pigeons, doves and hawks.

Impairment of fertility in an animal in accordance with the invention can take the form of either immunocontraception and immunosterilization. Immunosterilization means permanent, irreversible infertility, in contrast to immunocontraception wherein infertility is temporary or transient, and reversible. Immunocontraception and immunosterilization are both dependent on the antibody titer level in the serum of the subject, but immunosterilization is typically the result of ovarian pathology caused by the immune response of the animal, as evidenced by, for example, total destruction of the zona pellucida proteins and/or influx of leukocytes into the follicles. Reducing the number of boosters leads to reduced immune response which results in immunocontraception (i.e., infertility that is temporary and reversible) instead of immunosterilization.

Optionally, the zona pellucida protein can be conjugated to an immunogenic carrier protein. A conjugated form of an avian zona pellucida protein is suitable for use in embodiments of the fertility impairing vaccine containing an avian zona pellucida protein that is homologous with respect to the intended recipient; that is, where the avian zona pellucida protein is isolated from the same species to which it is to be administered. For example, where a

fertility impairing vaccine comprising chicken zona pellucida protein is intended for administration to chickens, it may be desirable to conjugate the chicken zona pellucida protein to an immunogenic carrier to augment the immune response of the recipient. However, it is to be understood that the invention includes  
5 administration of avian zona pellucida protein from any avian source to any bird, whether homologous or heterologous, and whether the avian zona pellucida protein is conjugated or unconjugated. For example, the invention includes administration of a fertility impairing vaccine comprising an unconjugated chicken zona pellucida to psittacine birds.

10 The fertility impairing vaccine is administered in a manner and an amount effective to cause the desired infertility in the animal. For example, to immunosterilize a dog or a cat using a protein vaccine of the invention, the vaccine is preferably administered in the form of a plurality of doses (typically about 1.0 mL for a dog, 0.5 mL for a cat), each dose containing avian zona  
15 pellucida protein or an immunogenic fragment thereof (or a mixed vaccine containing both avian and porcine zona pellucida proteins), in an amount of about 100  $\mu$ g to about 2 mg, more preferably about 100  $\mu$ g to about 400  $\mu$ g. To treat a bird, for example, a protein vaccine is preferably administered in the form of a plurality of doses (typically about 0.25 mL), each dose containing avian  
20 zona pellucida protein or an immunogenic fragment thereof (or a mixed vaccine containing both avian and porcine zona pellucida proteins), in an amount of about 10  $\mu$ g to about 2 mg, more preferably about 50  $\mu$ g to about 400  $\mu$ g.

A polynucleotide vaccine is preferably administered in one or more doses containing the plasmid, viral vector or naked polynucleotide in an amount  
25 of about 5  $\mu$ g to about 100  $\mu$ g. One of skill in the art can readily determine a suitable dosage for a particular animal, depending on the nature, size and overall health of the animal, as well as the condition to be treated.

For vaccines administered to mammals, an immunostimulant such as STDCM is typically present in a per dose amount of about 50  $\mu$ g to about 5  
30 mg, preferably in an amount of about 200  $\mu$ g to about 3.5 mg, more preferably in an amount of about 400  $\mu$ g to about 3.0 mg. For vaccines administered to birds, an immunostimulant such as STDCM is optionally present in a per dose amount of about 10  $\mu$ g to about 5 mg, preferably in an amount of about 50  $\mu$ g to about

3.5 mg, more preferably in an amount of about 1 mg to about 3 mg. When administered to a bird, the fertility impairing vaccine preferably contains ALOH as an adjuvant, or contains no adjuvant at all.

5 An initial injection of the vaccine is followed by one or more  
booster injections at two to four week intervals, although the boosters can be  
administered from about 9 days to about twelve months following the previous  
vaccination. Preferably, two booster shots are administered following the initial  
administration. The body's immunological response to the vaccine at this  
10 dosing regimen is expected to render the ovaries permanently inactive as a result  
of, for example, follicle disruption or destruction, as evidenced by  
immunocytochemical analysis and histological evaluation of the ovarian tissue  
of vaccinated subjects. Sterility is permanent and irreversible.  
Immunosterilization of animals in accordance with the present method is not  
expected to cause abnormal estrus cycles or other significant undesirable side  
15 effects in the vaccinated subjects.

When the fertility impairing vaccine is administered to a dog or a  
cat as described above, but with only one booster instead of two or more  
boosters, the vaccine is expected to result in immunocontraception (i.e.,  
temporary or transient, reversible infertility) rather than immunosterilization.  
20 Fertility impairment in dogs in accordance with the present method preferably is  
not expected to cause abnormal estrus cycles or other significant undesirable  
side effects in the vaccinated subjects.

Although the fertility impairing vaccine is typically administered  
by way of intramuscular injection, other forms of administration are also  
25 contemplated, including subcutaneous or intradermal administration, oral  
administration, as by food or water, topical administration, including  
transdermal administration, aerosol administration, cloacal or vaginal  
administration, intracoelomic administration, intranasal administration, and  
transconjunctival administration, including the use of eye drops. In addition,  
30 liposome-mediated, microsphere-mediated, and microencapsulation systems are  
all included as delivery vehicles for the fertility impairing vaccine of the present  
invention. The fertility vaccine can be formulated as a single dose vaccine and  
packaged for delivery through a dart or a bullet, such that it can be administered

to an animal from a distance. A single shot time-release delivery vehicle is also contemplated, wherein the avian zona pellucida protein or immunogenic fragment thereof (or the polynucleotide encoding the avian zona pellucida or immunogenic fragment thereof) is supplied in a single dose in two or more  
5 different forms, such as a free protein and a microencapsulated protein, to allow administration of the immunogenic agent to the animal over a longer period of time.

Advantages of the invention are illustrated by the following  
10 examples. However, the particular materials and amounts thereof recited in these examples, as well as other conditions and details, are to be interpreted to apply broadly in the art and should not be construed to unduly restrict or limit the invention in any way.

## 15 EXAMPLES

### Example I. Isolation of Avian Zona Pellucida Proteins

Perivitelline membranes were obtained from laid chicken eggs.  
20 Yolks were separated from whites by making a small hole in the egg and draining the albumin. The yolks were removed from the shell and the chalazae was removed and discarded. The perivitelline membranes (pvm) were either manually peeled away or punctured to drain the yolk, and the membranes were washed in sterile phosphate buffered saline solution. A tissue homogenizer  
25 (Powergam 700D) was used to homogenize the membranes. M. Wacławek et al. (Biol. Reprod., 59, 1230-1239 (1998)) reported a similar procedure for isolating perivitelline membranes from laid eggs that can also be used. Alternatively, perivitelline membranes can be isolated directly from ovarian follicles by preparing granulosa cell sheets substantially as described by A. Gilbert et al, J. Reprod. Fertil., 50, 179-181 (1977)).  
30

Purity was demonstrated and confirmed using one-dimensional and two-dimensional sodium dodecyl sulfate polyacrylamide gel electrophoresis combined with Western blot analysis, silver staining, and, at times, Coomassie

blue staining, using standard protocols. Proteins having molecular weights of 70 kD, 40 kD and 35 kD reacted with rabbit anti-pZP serum. Yolk and albumin controls did not react with the rabbit anti-pZP serum. The 70 kD protein was easily washed away from the perivitelline membrane. The 35 kD protein was approximately twice as abundant as the 40 kD protein and both had strong reactivity to the anti-pZP serum. One or both of these proteins are likely to be homologous to mammalian ZP3 (also known as ZPC), according to published reports (Y. Takeuchi et al., Eur. J. Biochem., 260, 736-742 (1999); M. Wacławek et al., Biol. Reprod., 59, 1230-1239 (1998)).

### **Example II. Preparation of Avian Zona Pellucida (aZP) Vaccine**

The vaccine is prepared by homogenizing a concentrated oily adjuvant concentrate with an aqueous antigen solution containing isolated aZP protein. The oily adjuvant concentrate contains a surfactant, lecithin, and an immunostimulant, synthetic trehalose dicorynomycolate (STDCM), in squalene oil. A typical adjuvant concentrate contains about 12% wt/vol (grams/100 mL) lecithin and about 25-50 mg/mL STDCM in squalene oil. The aqueous antigen solution contains the aZP protein preparation in saline or phosphate buffered saline (PBS) and Tween 80. When prepared for use in combination with an adjuvant concentrate to yield the vaccine composition, the aqueous composition typically contains 0.4% (vol/vol) Tween 80 and an amount of aZP calculated to yield a dose of about 100 µg to about 400 µg per vaccination. Vaccine doses for dogs are about 1 mL in volume.

Homogenizing is accomplished by combining adjuvant concentrate (to a final concentration of no greater than 10% vol/vol) with aqueous aZP solution and emulsifying using a Powergam 700D homogenizer at 15,000 rpm for 6 minutes. The resulting emulsion is then homogenized with phosphate buffered saline (PBS) (containing 0.4% vol/vol Tween 80) at 20,000 for 8-12 minutes. The homogenization process results in a vaccine composition that is an oil-in-water emulsion or possibly a water-in-oil-in-water emulsion. While the inventors do not intend that the invention be bound by any particular scientific theory, it is believed that the STDCM, an amphiphilic glycolipid, partitions to

the oil/water interfaces in the emulsion, and that the antigen is attracted to and associates with the STDCM at these interfaces.

### **Example III. Immunosterilization of Dogs using aZP Vaccine**

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Vaccinations. Female dogs are vaccinated with 200 µg of aZP per dose in a vaccine adjuvanted with synthetic trehalose dicorynomycolate (STDCM, commercially available from RIBI Immunochem Co., Hamilton, MT) in squalene oil. The amount of STDCM per dose was about 100 µg to about 2.5 mg. A 10X adjuvant concentrate as described in Example II is available from RIBI Immunochem Co., Hamilton, MT, and vaccines are prepared as described in Example II. Boosters (containing the same amount of aZP, 200 µg) are administered at 14-day intervals. Vaccinations are delivered to dogs intra-muscularly in the longissimus muscle (loin area). Booster injections are administered on the contra-lateral side.

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Antibody titers. Blood is drawn from each dog weekly, and serum antibody titers are determined using an enzyme linked immunosorbant assay (ELISA). Adjacent wells of a microwell plate are coated with 2 µg aZP, and incubated for 6 hours. The wells are then blocked with 5% bovine serum albumin (Sigma Chemical Co., St. Louis, MO) in TBST (Tris-buffered saline + 5% Tween-20) and incubated overnight. Wells are then loaded with the primary antibody (canine serum) in TBST at a 1:500 and 1:1,000 dilution and incubated for 4 hours. The wells are then washed and loaded with 50 µl of the secondary antibody (rabbit anti-dog IgG) and incubated for 2 hours. Color change is observed after the addition of *p*-nitrophenyl phosphate for 30 minutes and the reaction terminated by the addition of 3 M NaOH. The optical density is read at a 405-492 nm range on a Spectramax spectrophotometer. The dog's pre-immune serum serves as the negative control.

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**Example IV. Administration of Mixed Chicken/Porcine ZP Vaccine  
to Psittacines**

Vaccinations. Chickens are vaccinated with 200 µg of mixed zona  
5 pellucida protein (1:1, aZP:pZP) per dose (1 mL volumes) in a vaccine  
adjuvanted with Freund's complete adjuvant. The birds are vaccinated with  
three injections administered at approximately two week intervals. Under  
veterinary supervision, vaccinations are delivered to chickens intramuscularly in  
the deep pectoral muscle. Booster injections are administered on the  
10 contra-lateral side.

Antibody titers. Blood is drawn from each bird at the time of each of the  
three injections, and about three weeks following the last injection. Serum  
antibody titers (IgG) are determined using an enzyme linked immunosorbant  
assay (ELISA). Adjacent wells of a microwell plate are coated with 2 µg aZP,  
15 and incubated for 6 hours. The wells are then blocked with 5% bovine serum  
albumin (Sigma Chemical Co., St. Louis, MO) in TBST (Tris-buffered saline +  
5% Tween-20) and incubated overnight. Wells are then loaded with the primary  
antibody (i.e., avian serum) in TBST at a 1:500 and 1:1,000 dilution and  
incubated for 4 hours. The wells are then washed and loaded with 50 µl of the  
20 secondary antibody (rabbit anti-chicken IgG) and incubated for 2 hours. Color  
change is observed after the addition of *p*-nitrophenyl phosphate for 30 minutes  
and the reaction is terminated by the addition of 3 M NaOH. The optical density  
is read at a 405-492 nm range on a Spectramax spectrophotometer. The  
chickens' pre-immune serum serves as the negative control.

Egg laying. Eggs are counted beginning on the day of the first  
25 injection to evaluate reduction in egg production, providing direct evidence of  
fertility impairment.

The complete disclosure of all patents, patent applications, and  
30 publications cited herein are incorporated by reference. The foregoing detailed  
description and examples have been given for clarity of understanding only. No  
unnecessary limitations are to be understood therefrom. The invention is not

limited to the exact details shown and described, for variations obvious to one skilled in the art will be included within the invention defined by the claims.



**WHAT IS CLAIMED IS:**

1. A fertility impairing vaccine comprising at least one component selected from the group consisting of (a) a polypeptide comprising an avian zona pellucida protein or an immunogenic fragment thereof and (b) a polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising an avian zona pellucida protein or an immunogenic fragment thereof.
2. The fertility impairing vaccine of claim 1 wherein the avian zona pellucida protein is a glycoprotein.
3. The fertility impairing vaccine of claim 1 wherein the avian zona pellucida protein is a naturally occurring protein.
4. The fertility impairing vaccine of claim 1 wherein the avian zona pellucida protein is a recombinant protein.
5. The fertility impairing vaccine of claim 1 wherein the avian zona pellucida protein is a chemically or enzymatically synthesized protein.
6. The fertility impairing vaccine of claim 1 further comprising an immunological adjuvant.
7. The fertility impairing vaccine of claim 6 wherein the immunological adjuvant comprises synthetic trehalose dicorynomycolate.
8. The fertility impairing vaccine of claim 1 further comprising squalene oil.
9. The fertility impairing vaccine of claim 1 wherein the polypeptide further comprises at least one epitope selected from the group consisting of a T cell epitope, a helper T cell epitope and a B cell epitope.

10. The fertility impairing vaccine of claim 1 which is an immunosterilant vaccine.
11. The fertility impairing vaccine of claim 1 which is an immunocontraceptive vaccine.
12. The fertility impairing vaccine of claim 1 wherein the polynucleotide comprises a vector.
13. The fertility impairing vaccine of claim 12 wherein the vector is a plasmid.
14. The fertility impairing vaccine of claim 12 wherein the vector is a viral vector.
15. The fertility impairing vaccine of claim 1 wherein the polynucleotide further comprises a regulatory sequence operably linked to the nucleotide sequence encoding the polypeptide comprising the avian zona pellucida protein or immunogenic fragment thereof.
16. The fertility impairing vaccine of claim 1 wherein the polynucleotide further comprises an immunostimulatory sequence.
17. A method for impairing the fertility of an animal comprising administering to the animal a fertility impairing vaccine of any of the preceding claims wherein the vaccine is administered in a manner and an amount effective to cause fertility impairment in the animal.
18. The method of claim 17 wherein the vaccine causes temporary, reversible infertility in the animal.
19. The method of claim 17 wherein the vaccine causes permanent, irreversible infertility in the animal.

20. The method of claim 17 wherein the animal is a mammal.
21. The method of claim 20 wherein the mammal is selected from the group consisting a horse, a deer, an elephant, a rat, a mouse, a rabbit, a ferret, a dog and a cat.
22. The method of claim 21 wherein the animal is a dog.
23. The method of claim 21 wherein the animal is a cat.
24. The method of claim 17 wherein the animal is a bird.
25. The method of claims 17-24 wherein the fertility impairing vaccine further comprises at least one component selected from the group consisting of (a) a polypeptide comprising a porcine zona pellucida protein or an immunogenic fragment thereof and (b) a polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising a porcine zona pellucida protein or an immunogenic fragment thereof.
26. The method of claim 25 wherein the fertility impairing vaccine is a polypeptide vaccine, and wherein the ratio (aZP:pZP) of avian zona pellucida protein (aZP) to porcine zona pellucida protein (pZP) is about 100:1 to about 1:100.
27. The fertility impairing vaccine of claims 1-16 further comprising at least one component selected from the group consisting of (a) a polypeptide comprising a porcine zona pellucida protein or an immunogenic fragment thereof and (b) a polynucleotide comprising a nucleotide sequence encoding a polypeptide comprising a porcine zona pellucida protein or an immunogenic fragment thereof.

28. The fertility impairing vaccine of claim 27 which is a polypeptide vaccine wherein the ratio (aZP:pZP) of avian zona pellucida protein (aZP) to porcine zona pellucida protein (pZP) is about 100:1 to about 1:100.

**DECLARATION**

We, Richard Fayrer-Hosken and Branson W. Ritchie, declare that: (1) our respective citizenships and residence/mailling addresses are indicated below; (2) we have reviewed and understand the contents of the specification identified below, including the claims, as amended by any amendment specifically referred to herein, (3) we believe that we are the original, first, and joint inventors of the subject matter in

**FERTILITY IMPAIRING VACCINE CONTAINING AVIAN ZONA PELLUCIDA  
PROTEIN AND METHOD OF USE**

Filing Date: December 28, 2001

Serial No.: Unknown

described and claimed therein and for which a patent is sought; and (4) we hereby acknowledge our duty to disclose to the United States Patent and Trademark Office all information known to us to be material to the patentability as defined in Title 37, Code of Federal Regulations, §1.56.\*

We hereby claim foreign priority benefits under Title 35, United States Code, §119(a)-(d) or §365(b) of any foreign application(s) for patent or inventor's certificate listed below, or §365(a) of any PCT international application which designates at least one country other than the United States of America listed below, and have also identified below any foreign application for patent or inventor's certificate, or any PCT international application having a filing date before that of the application on the basis of which priority is claimed:

- a. ☒ no such applications have been filed.  
b. ☐ such applications have been filed as follows:

<b>FOREIGN APPLICATION(S), IF ANY, CLAIMING PRIORITY UNDER 35 USC §119(a)-(d), §365(a), and/or §365(b)</b>			
<b>COUNTRY</b>	<b>APPLICATION NUMBER</b>	<b>DATE OF FILING (day, month, year)</b>	<b>DATE OF ISSUE (day, month, year)</b>

<b>ALL FOREIGN APPLICATIONS, IF ANY, FILED BEFORE THE PRIORITY APPLICATION(S)</b>			
<b>COUNTRY</b>	<b>APPLICATION NUMBER</b>	<b>DATE OF FILING (day, month, year)</b>	<b>DATE OF ISSUE (day, month, year)</b>

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\* Title 37, Code of Federal Regulations, §1.56 is reproduced on the attached page.

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*Declaration*

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*Applicants: Fayrer-Hosken et al.*

*Serial No.: Unknown*

*Filing Date: December 28, 2001*

*Title: FERTILITY IMPAIRING VACCINE CONTAINING AVIAN ZONA PELLUCIDA PROTEIN AND METHOD OF USE*

We hereby claim the benefit under Title 35, United States Code §119(e) of any United States provisional application(s) listed below.

- a. ☐ no such applications have been filed.  
b. ☒ such applications have been filed as follows:

PROVISIONAL APPLICATION(S), IF ANY, UNDER 35 USC §119(e)	
APPLICATION NUMBER	DATE OF FILING (day, month, year)
60/141,929	1 July 1999
60/162,984	2 November 1999

We hereby claim the benefit under Title 35, United States Code, §120 of any United States applications or §365(c) of any PCT international application(s) designating the United States of America, listed below.

- a. ☐ no such applications have been filed.  
b. ☒ such applications have been filed as follows:

APPLICATION NUMBER	DATE OF FILING (day, month, year)	STATUS (patented, pending, abandoned)
PCT/US00/18051	30 June 2000	Pending

Insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT international application in the manner provided by the first paragraph of Title 35, United States Code, §112, we acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, §1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of this application.

The undersigned declare further that all statements made herein of their own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Declaration

Page 2 of 4

Applicants: Fayrer-Hosken et al.

Serial No.: Unknown

Filing Date: December 28, 2001

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Declaration

Page 3 of 4

Applicants: Fayrer-Hosken et al.

Serial No.: Unknown

Filing Date: December 28, 2001

Title: FERTILITY IMPAIRING VACCINE CONTAINING AVIAN ZONA PELLUCIDA PROTEIN AND METHOD OF USE

Wherefore, we pray that Letters Patent be granted to us for the invention described and claimed in the specification identified above and we hereby subscribe our names to the foregoing specification, claims, and Declaration, on the date indicated below.

Name: Richard Fayrer-Hosken Date  
Citizenship: United Kingdom  
Residence: P.O. Box 27, Winterville, Georgia 30683  
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2001  
Name: Branson W. Ritchie Date  
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Declaration

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Applicants: Fayrer-Hosken et al.

Serial No.: Unknown

Filing Date: December 28, 2001

Title: FERTILITY IMPAIRING VACCINE CONTAINING AVIAN ZONA PELLUCIDA PROTEIN AND METHOD OF USE

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100  
1/22/02  
Name: Richard Fayrer-Hosken Date  
Citizenship: United Kingdom  
Residence: P.O. Box 27, Winterville, Georgia 30683  
Mailing Address: Same as above  
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Name: Branson W. Ritchie Date  
Citizenship: United States of America  
Residence: 1080 Barnett Place, Athens, Georgia 30605  
Mailing Address: Same as above  
(If different than Residence)

Cynthia G. Juff

Notary Public, Athens-Clarke County, Georgia  
My Commission Expires July 13, 2002

*Declaration*

Page 4 of 4

*Applicants: Fayrer-Hosken et al.*

*Serial No.: Unknown*

*Filing Date: December 28, 2001*

*Title: FERTILITY IMPAIRING VACCINE CONTAINING AVIAN ZONA PELLUCIDA PROTEIN AND METHOD OF USE*

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§ 1.56 Duty to disclose information material to patentability.

(a) A patent by its very nature is affected with a public interest. The public interest is best served, and the most effective patent examination occurs when, at the time an application is being examined, the Office is aware of and evaluates the teachings of all information material to patentability. Each individual associated with the filing and prosecution of a patent application has a duty of candor and good faith in dealing with the Office, which includes a duty to disclose to the Office all information known to that individual to be material to patentability as defined in this section. The duty to disclose information exists with respect to each pending claim until the claim is cancelled or withdrawn from consideration, or the application becomes abandoned. Information material to the patentability of a claim that is cancelled or withdrawn from consideration need not be submitted if the information is not material to the patentability of any claim remaining under consideration in the application. There is no duty to submit information which is not material to the patentability of any existing claim. The duty to disclose all information known to be material to patentability is deemed to be satisfied if all information known to be material to patentability of any claim issued in a patent was cited by the Office or submitted to the Office in the manner prescribed by §§1.97(b)-(d) and 1.98. However, no patent will be granted on an application in connection with which fraud on the Office was practiced or attempted or the duty of disclosure was violated through bad faith or intentional misconduct. The Office encourages applicants to carefully examine:

- (1) Prior art cited in search reports of a foreign patent office in a counterpart application, and
- (2) The closest information over which individuals associated with the filing or prosecution of a patent application believe any pending claim patentably defines, to make sure that any material information contained therein is disclosed to the Office.

(b) Under this section, information is material to patentability when it is not cumulative to information already of record or being made of record in the application, and

- (1) It establishes, by itself or in combination with other information, a prima facie case of unpatentability of a claim; or
- (2) It refutes, or is inconsistent with, a position the applicant takes in:
  - (i) Opposing an argument of unpatentability relied on by the Office, or
  - (ii) Asserting an argument of patentability.

A prima facie case of unpatentability is established when the information compels a conclusion that a claim is unpatentable under the preponderance of evidence, burden-of-proof standard, giving each term in the claim its broadest reasonable construction consistent with the specification, and before any consideration is given to evidence which may be submitted in an attempt to establish a contrary conclusion of patentability.

(c) Individuals associated with the filing or prosecution of a patent application within the meaning of this section are:

- (1) Each inventor named in the application;
- (2) Each attorney or agent who prepares or prosecutes the application; and
- (3) Every other person who is substantively involved in the preparation or prosecution of the application and who is associated with the inventor, with the assignee or with anyone to whom there is an obligation to assign the application.

(d) Individuals other than the attorney, agent or inventor may comply with this section by disclosing information to the attorney, agent, or inventor.

(e) In any continuation-in-part application, the duty under this section includes the duty to disclose to the

Office all information known to the person to be material to patentability, as defined in paragraph (b) of this section, which became available between the filing date of the prior application and the national or PCT international filing date of the continuation-in-part application.

*Declaration*

Page 4 of 4

*Applicants: Fayrer-Hosken et al.*

*Serial No.: Unknown*

*Filing Date: December 28, 2001*

*Title: FERTILITY IMPAIRING VACCINE CONTAINING AVIAN ZONA PELLUCIDA PROTEIN AND METHOD OF USE*

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